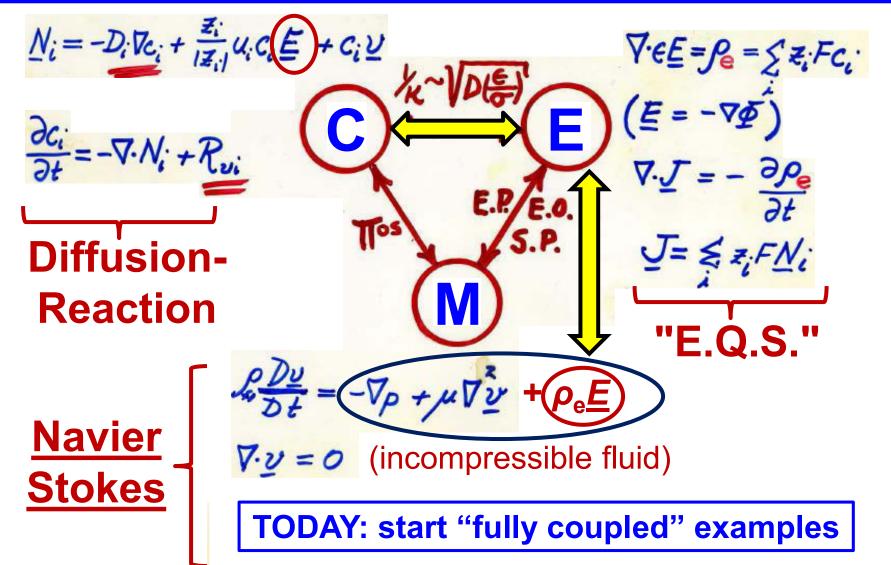
FFF: Complete Description of Coupled Transport and Biomolecular Interactions



Term Paper Project

Enzymatic Targeting of the Stroma Ablates Physical Barriers to Treatment of Pancreatic Ductal Adenocarcinoma

Cancer Cell 2012

2013

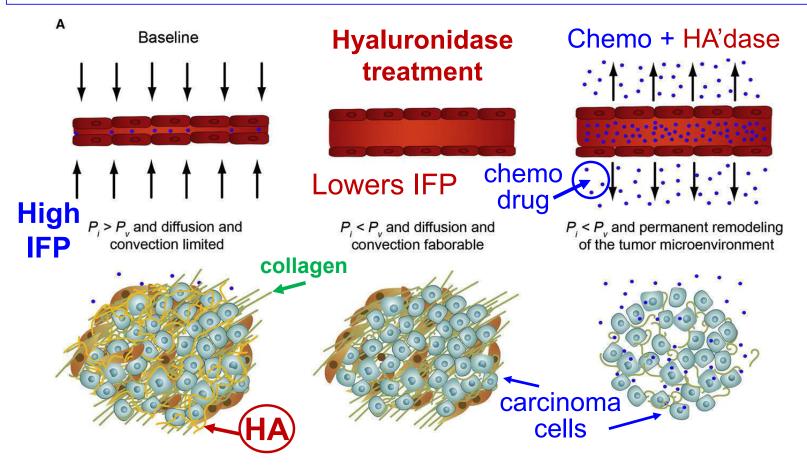
Paolo P. Provenzano,¹ Carlos Cuevas,⁴ Amy E. Chang,¹ Vikas K. Goel,¹ Daniel D. Von Hoff,⁵ and Sunil R. Hingorani^{1,2,3,*} ¹Clinical Research Division ²Public Health Sciences Division Fred Hutchinson Cancer Research Center, Seattle, WA 98109, USA ³Division of Medical Oncology, University of Washington School of Medicine, Seattle, WA 98195, USA ⁴Department of Radiology, University of Washington, Seattle, WA 98195, USA ⁵Clinical Translational Research Division, Translational Genomics Research Institute, Scottsdale, AZ 85259, USA ^{*}Correspondence: srh@fhcrc.org DOI 10.1016/j.ccr.2012.01.007

Hyaluronan, fluid pressure, and stromal resistance in pancreas cancer British J of Cancer

P P Provenzano^{1,4} and S R Hingorani^{*,1,2,3}

¹Clinical Research Division, Fred Hutchinson Cancer Research Center, Seattle, WA 98109, USA; ²Public Health Sciences Division, Fred Hutchinson Cancer Research Center, Seattle, WA 98109, USA; ³Division of Medical Oncology, University of Washington School of Medicine, Seattle, WA 98195, USA

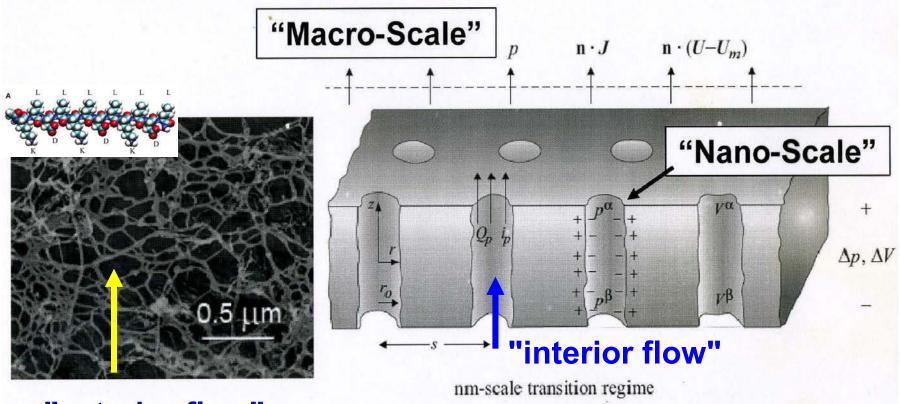
Fig. 7: Altering Physicomechanics & Remodeling Stroma in Pancreatic Ductal Adenocarcinoma to Therapeutic Advantage



Courtesy of Elsevier, Inc., http://www.sciencedirect.com. Used with permission. Source: Provenzano, Paolo P. et al. "Enzymatic targeting of the stroma ablates physical barriers to treatment of pancreatic ductal adenocarcinoma." Cancer Cell 21, no. 3 (2012): 418-429.

(A) High Interstitial Fluid Pressure → impedes diffusion & convection of chemo drugs.
 (B) Enzymatic degradation of stromal HA decreases IFP and relieves physical constraints on small molecule perfusion. (C) Combined enzymatic + cytotoxic therapy permanently remodels the tumor microenvironment to favor drug delivery

Fluid Flow in and across "Bio Porous Media: Tissues, Gels, Intra- and Extra-cellular space

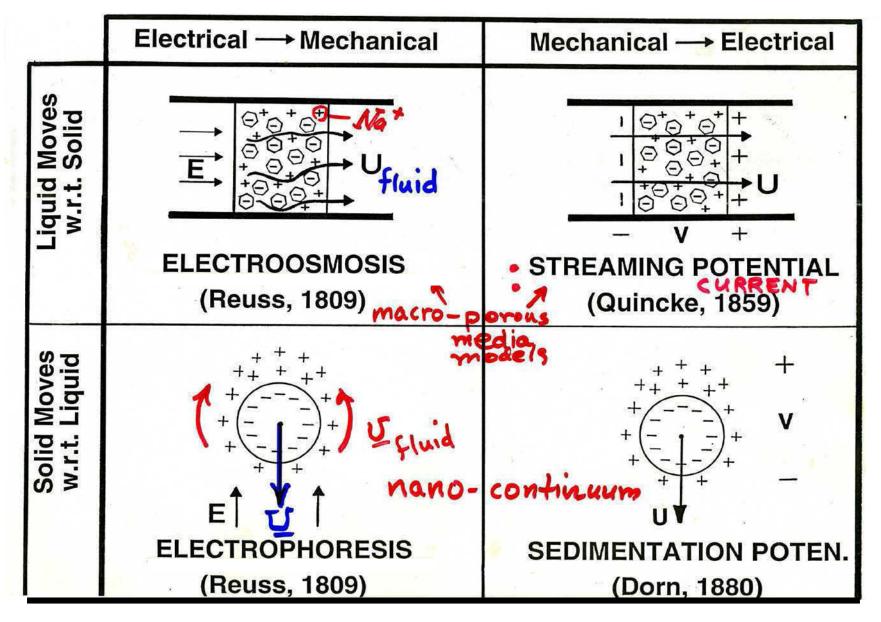


"exterior flow"

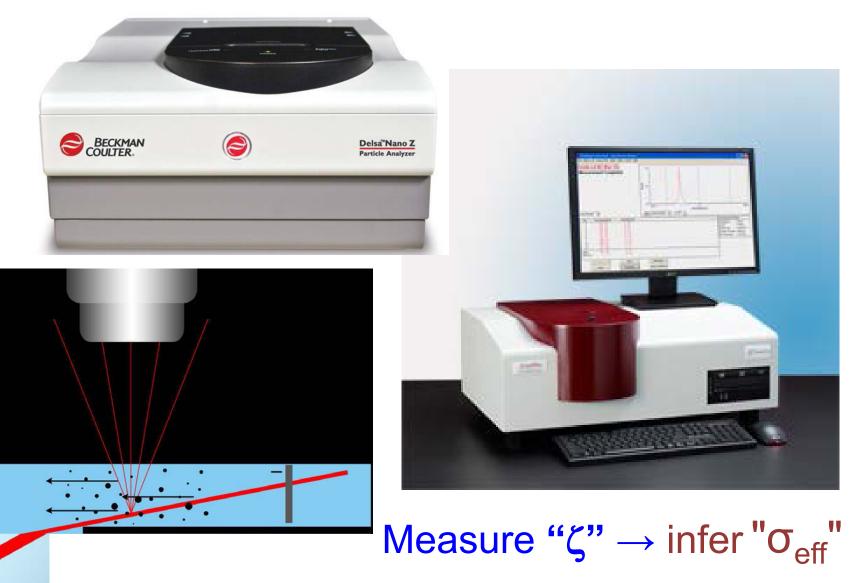
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Local "nano"interior / exterior flows vs. "macro" Darcy model

ELECTROKINETIC PHENOMENA



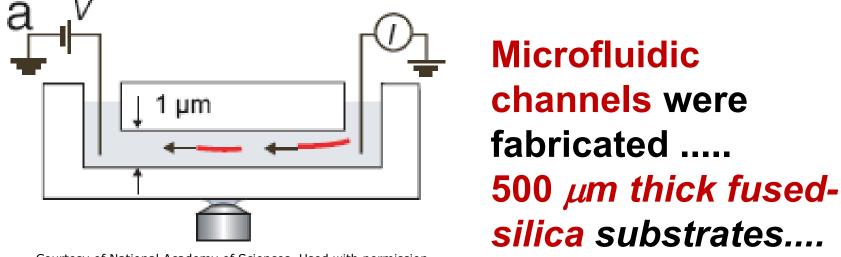
Zeta Potential (particle charge) Instruments



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Electrophoresis of individual microtubules in microchannels PNAS 2007

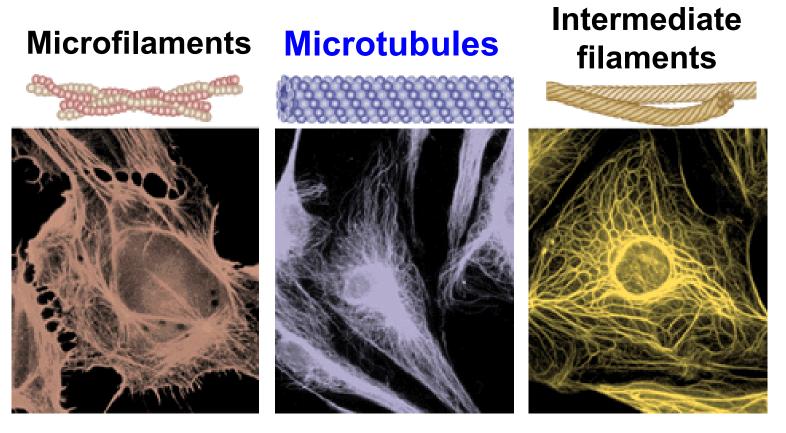
M. G. L. van den Heuvel, M. P. de Graaff, S. G. Lemay, and C. Dekker*



Courtesy of National Academy of Sciences. Used with permission. Source: Van den Heuvel, M. G. L. et al. "Electrophoresis of individual microtubules in microchannels." Proceedings of the National Academy of Sciences 104, no. 19 (2007): 7770-7775.

Microfabricated slit-like fluidic channels form an excellent system to confine and observe the electrophoretic motion of individual fluorescently labeled biomolecules, such as microtubules, actin filaments, or virus particles.

Primary Structural Filaments of the Cytoskeleton

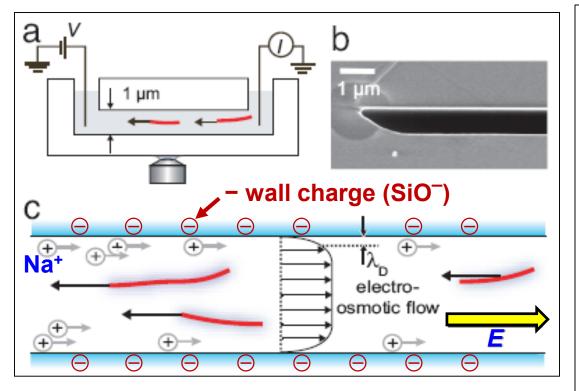


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Electrophoresis of individual microtubules in microchannels PNAS 2007

M. G. L. van den Heuvel, M. P. de Graaff, S. G. Lemay, and C. Dekker*

Kavli Institute of Nanoscience, Delft University of Technology, Lorentzweg 1, 2628 CJ, Delft, The Netherlands



Courtesy of National Academy of Sciences. Used with permission. Source: Van den Heuvel, M. G. L. et al. "Electrophoresis of individual microtubules in microchannels." Proceedings of the National Academy of Sciences 104, no. 19 (2007): 7770-7775. The electrophoretic mobility of molecules is a fundamental property.... In ensemble measurements, such as gel electrophoresis or dynamic light scattering, the differences between individual molecules are obscured. Here, individual microtubules are visible by fluorescent labeling, and their electrophoretic motion can be imaged using fluorescence microscopy

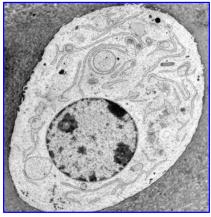
2013 Ø PLOS

Cytoplasmic Electric Fields and Electroosmosis: Possible Solution for the Paradoxes of the Intracellular Transport of Biomolecules

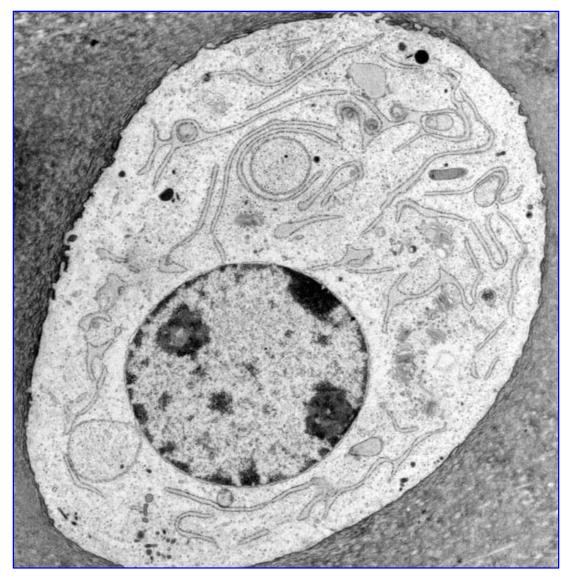
Victor P. Andreev^{1,2,3}*

Abstract

Electroosmotic flow might play an important role in the intracellular transport of biomolecules. The paper presents two mathematical models describing the role of electroosmosis in the transport of negatively charged messenger proteins to the negatively charged nucleus and in the recovery of the fluorescence after photobleaching

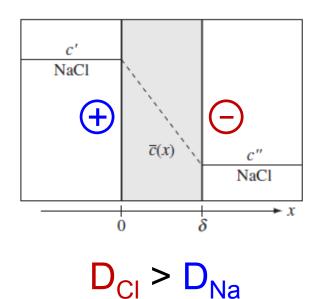


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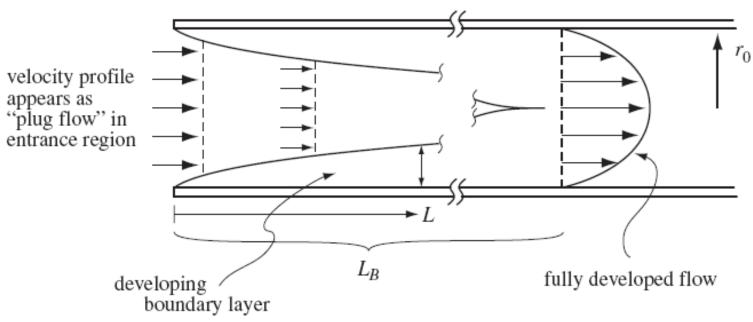


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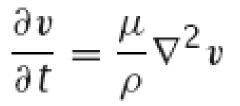
Transmembrane potentials everywhere inside (ion channels + diffusion potentials)



Transition to Fully Developed Flow in Channel



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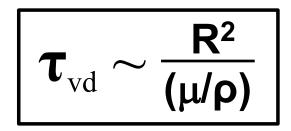
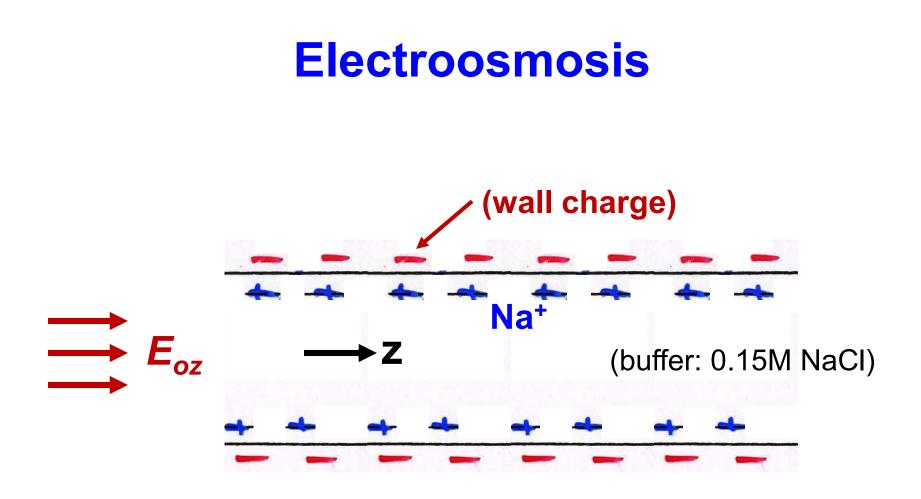
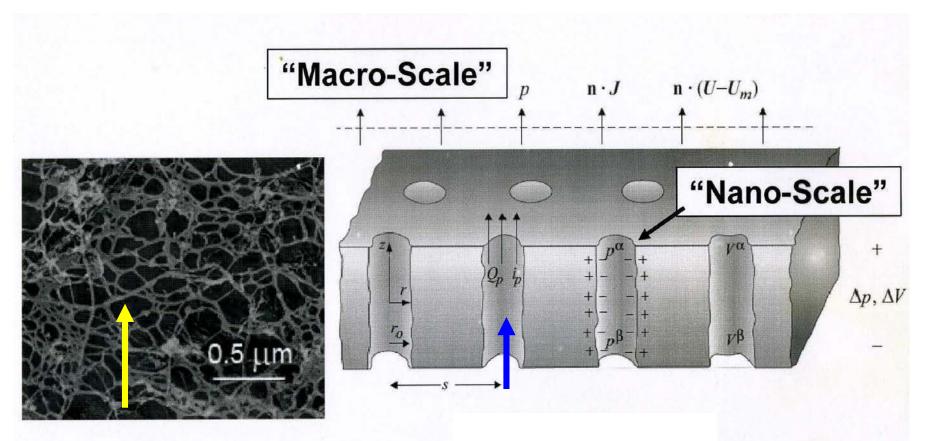


Table B.4
p. 294
$$\begin{array}{c} \left[\underbrace{v} \cdot \nabla \underbrace{v} \right] = \underbrace{v}_{r} \frac{\partial v_{e}}{\partial x} + \underbrace{v}_{e} \frac{\partial v_{e}}{\partial x} + \underbrace{v}_{e} \frac{\partial v_{e}}{\partial z} \\ \hline \underbrace{z}_{e} \circ \frac{\partial v}{\partial r} + \underbrace{v}_{e} \frac{\partial v_{e}}{\partial z} + \underbrace{v}_{e} \frac{\partial v_{e}}{\partial z} \\ (cylindrica(coord)) \circ \end{array}$$

$$\begin{array}{c} & \left(cylindrica(coord) \circ \right) \\ & & \left(cylind$$



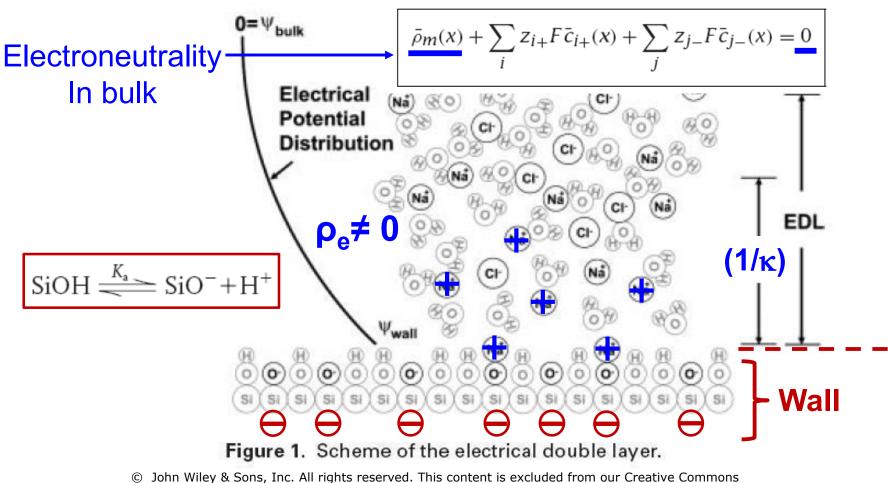
Charged fibers and "pores" in Bio-porous Media: Tissues, Gels, Intra- and Extra-cellular space



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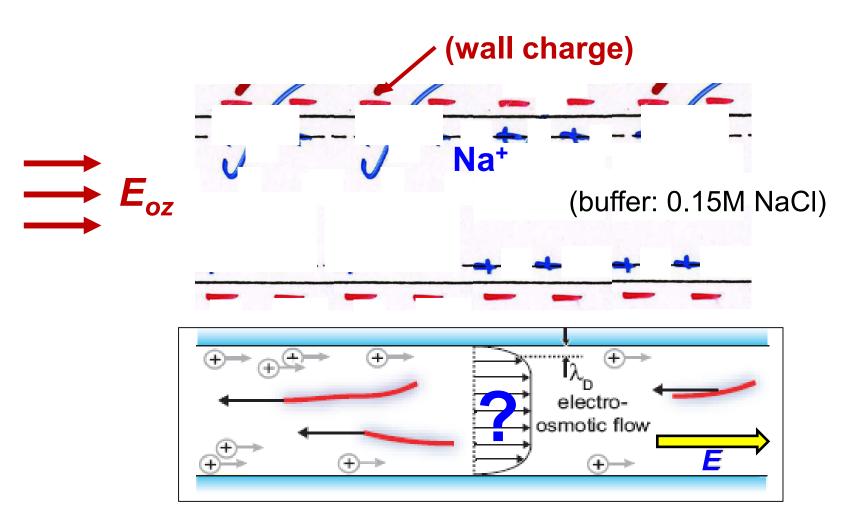
Tandon, Kirby et al. Electrophoresis 2008, 29, 1092-1101

Zeta potential and electroosmotic mobility in microfluidic devices fabricated from hydrophobic polymers: The origins of charge

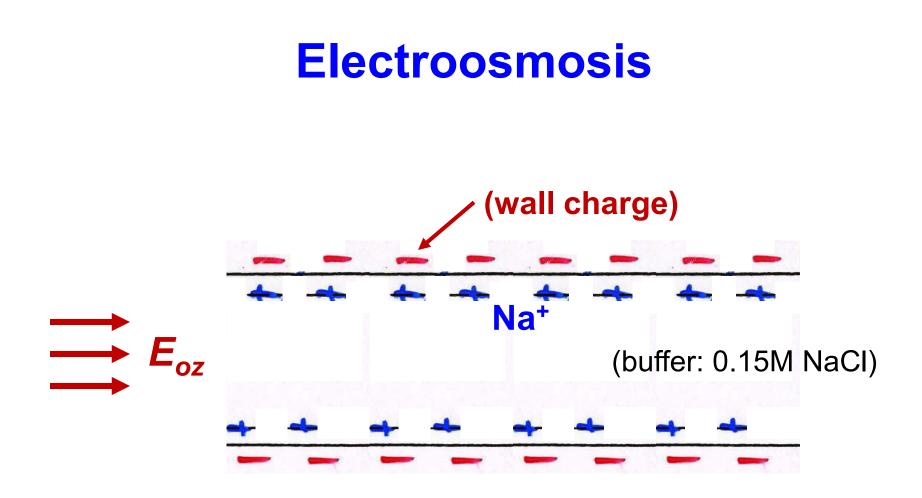


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Electroosmosis



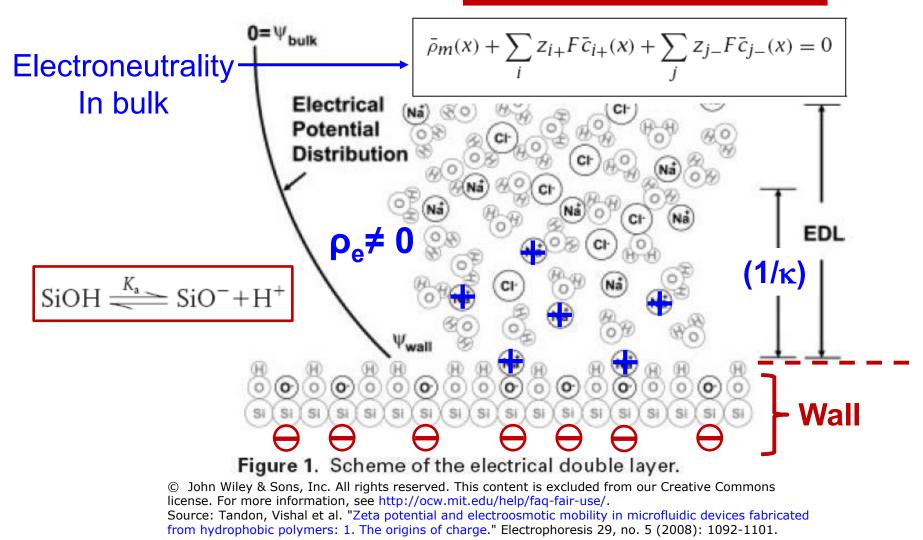
Courtesy of National Academy of Sciences. Used with permission. Source: Van den Heuvel, M. G. L. et al. "Electrophoresis of individual microtubules in microchannels." Proceedings of the National Academy of Sciences 104, no. 19 (2007): 7770-7775.



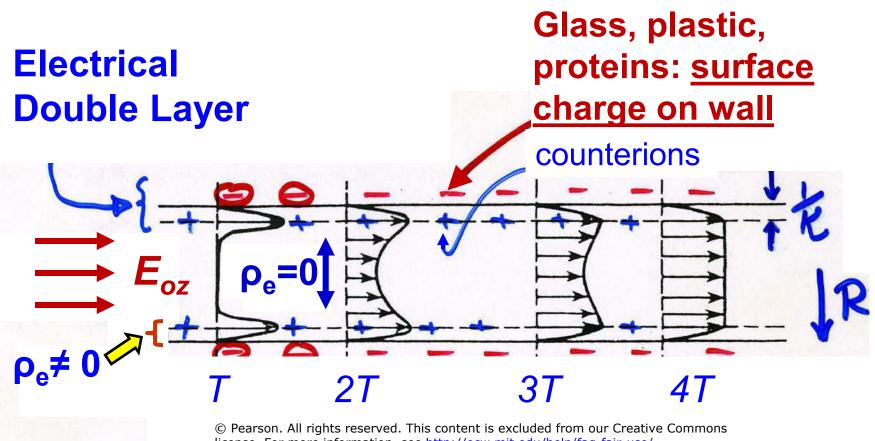
Where is the $\rho_{e}E$ force on the fluid??

Tandon, Kirby et al. Electrophoresis 2008, 29, 1092–1101

Zeta potential and electroosmotic mobility in microfluidic devices fabricated from hydrophobic polymers: The origins of charge

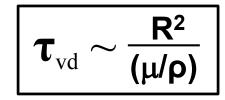


Electroosmosis: Turn-on Transient



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 $-\nabla p + \mu \nabla^2 v + \rho_e \boldsymbol{E} = 0$



Superposition

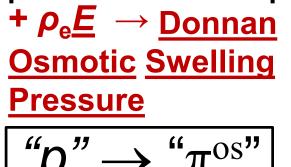
Uz(r) -?? 0=~V (Electroosmosis) Stokes \rightarrow (Poiseuille) ╋

LAWS (1) $N_i = -D_i \nabla c_i + \frac{z_i}{|z_i|} u_i c_i E + c_i \mathcal{V}$ $(2)(\partial c_i/\partial t) = -\nabla N_i + R_i$ (3) $\nabla \cdot e E = e = S = S = Fc;$ $(4) E^{Tor} = -\nabla \overline{\phi}^{Tor}$ $(\mathbf{E})_{(5)} \nabla \mathbf{J} = -(\partial \mathbf{e}/\partial t)$ $(6) J = \sigma E^{T + r} e^{\gamma} + () \nabla c_i = \sum z_i F N_i$ $(7) \mathcal{D}_{\underline{\mu}} = (-\nabla p + \mu \nabla u + \rho \underline{E}) \simeq 0$ $(8) \nabla \upsilon = 0$ C- ; N+,N-; E;ど (+) EQNS. IN 14 unknowns !!

LAWS (1) $\underline{N}_{i} = -D_{i}\nabla c_{i} + \frac{\overline{z}_{i}}{|\overline{z}_{i}|}u_{i}c_{i}\underline{E} + c_{i}\underline{v}$ $(2)(\partial c_i/\partial t) = -\nabla N_i + R_i$ (3) $\nabla \cdot e \underline{E} = e = \sum_{i=1}^{Tot} E = \sum_{i=1}^{Tot} E_i E_i$ $(4) E^{Tot} = -\nabla \phi^{Tot}$ $(5) \nabla J = -(\partial p/\partial t)$ (6) $J = \sigma E^{T+} \rho v + () \nabla c_i$ (7) <u>Du</u> =

Initial equilibrium (t < 0):

Tissue, tumor, can SWELL due to electrostatic repulsive interactions in ECM



20.430J / 2.795J / 6.561J / 10.539J Fields, Forces, and Flows in Biological Systems $\mathsf{Fall}\ 2015$

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