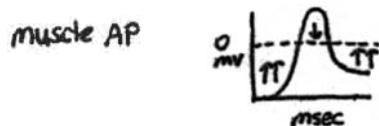


Lecture notes courtesy of Wyan-Ching Mimi Lee. Used with permission.

3/3/04



reversal potential = 0 (or -15 mV, depending on muscle cell)

b/c AChR lets in  $\text{Na}^+$  and  $\text{K}^+$ 

- collagenase to get postsynaptic side clear, patch clamp (w/ ACh in blunt microelectrode), look at current flow (magnitude + direction)

↳ voltage clamp

 $(V_m < 0, \text{ current negative} = \text{inward})$  $(V_m > 0, \text{ current positive} = \text{outward})$ 

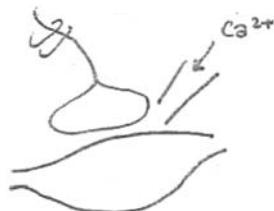
- ionotropic receptors have reversal potentials

- if  $E_{\text{reversal}}$  above or below threshold, determines if excitatory or inhibitory
- if  $E_{\text{reversal}} = \text{threshold}$ , will not affect either way

- stimulate extracellularly presynaptic, record intracellularly postsynaptic

- need  $\text{Ca}^{2+}$  ( $\downarrow$ ) for presynaptic transmitter release, EPSP

- cobalt interferes w/ natural  $\text{Ca}^{2+}$  leak from slightly damaged muscle



$\text{Ca}^{2+}$  must be near synaptic cleft, present around when AP reaches terminal (or right before reaches terminal)

treated whole system w/ TTX, increased stimulus intensity to passively conduct to terminal:

found that pure voltage could mimic AP : evidence for voltage-gated  $\text{Ca}^{2+}$  channels

- intracellular presynaptic injection of  $\text{Ca}^{2+}$  also gives EPSP

- minis - correspond to small releases of ACh (b/c can affect w/ curare, neostigmine)

- make up most EPSP: reduce  $\text{Ca}^{2+}$ , stimulate presynaptically, record postsynaptically
  - effect comes in quantal sizes ( $J/\text{Hz}$ )

- Gaussian distribution around 0.4 mV

↳ direct evidence for quantal transmission

- large number of vesicles w/ equal  $\tau_p$ , independent probability of release (from Poisson curve assumptions)

- make preparations of synapses, synaptic vesicles
  - synaptobrevin - V-SNARE (important in exocytosis)
    - target of botulinum toxin (BOTOX) & tetanus toxin
    - mutants homologous to yeast mutants w/ exocytic defect
      - yeast cycle vesicles between inner & outer lamellae
      - bad cycling due to mutant v-snakes & t-snakes
      - vesicle endocytosis looked at in cell-free dog pancreas system; these same proteins showed up
  - synaptotagmin -  $\text{Ca}^{2+}$  binding domain: this is the calcium sensor for vesicle exocytosis
  - neuromuscular junctions can be modulated
    - (eg by chip screaming - potentiated by adrenaline)
    - potentiated by sympathetic stimulation (neuromuscular transmission)
      - Orbelli effect (controversy: presynaptic or postsynaptic?): turns out is both
        - do quantal analysis before + after sympathetic stimulation: more quanta released per action potential will give more double, triple, etc releases (average # releases), if presynaptic effect; if postsynaptic (eg more AChRs, or more sensitive AChRs, will not affect placement of peaks by shifting histogram right (eg 0.4 mV  $\rightarrow$  0.6 mV)
        - $P(x) = e^{-m} \frac{m^x}{x!}$  (but there is better way)
        - $P_0 = e^{-m} \left( \frac{m^{x+1}}{(x+1)!} \right)$  } easy to tell 0's from 1's :  $P_0 = e^{-m} \left( \frac{m^{x+1}}{(x+1)!} \right)$
        - so: if presynaptic  $\downarrow$  (increase mean quantal content) ( $P_0$  decreases)
        - if postsynaptic measure fraction of failures, see if goes up or down
        - if postsynaptic, will affect quantal size
    - mean quantal content = average # quanta (vesicles) released per stimulus (not contents of vesicle)
      - measure fraction of failures, plug into  $P_0 = e^{-m}$  no dimensions
      - $\downarrow$  fraction of failures
      - higher quanta release # = lower probability of failure

to measure change in quantal size, look at minis, look at size of mEPSPs

- this will increase if Orbelli effect postsynaptic

- quantal size  $V_i$  = average voltage displacement of quantum of spontaneous miniature potential  
 $\underline{\underline{\text{or}}}$   
not physical size of vesicle (all ~ same size, same density of transmitter)

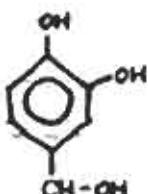
### norepinephrine

Orbelli effect: more adrenaline comes down sympathetic axons to neuromuscular junctions

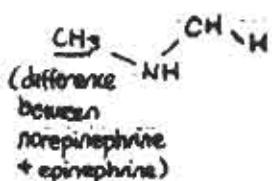
adrenal gland secretes another compound (cycles around your body): norepinephrine?

- there result in presynaptic enhanced quantal release. (NE)

- also postsynaptic response from circulating hormone (EPI)



phenylalanine → tyrosine → tyramine → dopamine → norepinephrine  
 adrenaline ← (norepinephrine)  
 ↘ NE



adrenaline from adrenal medulla (on top of kidney)

- epinephrine same in Greek

↳ EPI

norepinephrine & epinephrine both from phenylalanine, differ by methyl group

### $\beta$ - propanolol

antagonist: isoproterenol (EPI antagonist) drug used for asthma in hospitals, stage fright

heart conditions

- applying this gives no change in quantal size

- better music performance

epinephrine - agonist = isoproterenol

(adrenaline) antagonist =  $\beta$  - propanolol (blocks adrenergic response on postsynaptic side)

presynaptic - look at number of failures (Cuba found less failures)

norepinephrine - agonist = norepinephrine

(noradrenaline) antagonist = clonidine (blocks presynaptic response, gives pure postsynaptic)

postsynaptic - look at quantal size (will be bigger w/ Orbelli effect)

- postsynaptic so increase actually from

epinephrine closing  
leakage channels

depolarizing current leaks  
out less if R↑ (Rinput)

$$V = I R$$

so R↑, VT

depolarizes more & longer



3

I from ionotropic AChRs: doesn't change, depolarizes cell, but leakage out b/c muscle is big; plug up leakage holes in muscle, increase R

- modulated synapses - have volume control
- norepinephrine + epinephrine are modulatory transmitters (gain control)

7.29J / 9.09J Cellular Neurobiology

Spring 2012

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