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Polyketide Biosynthesis

Type II Polyketide Biosynthesis: Aromatic Polyketides

**Aromatic polyketides were at the start of the biosynthetic field
One of the first:**

- **biomimetic synthesis (Collie, 1907)**
- **isotopic labeling studies (Birch, 1955)**
- **gene cluster sequence (Hopwood, 1984)**

Figures removed due to copyright reasons.

Please see: Fig.2A-D in *Top Curr Chem* 209 (2000): 1-51.

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We now know that there are many variations of the polyketide pathway

Type I PKS: Catalytic domains are linked together on same polypeptide

Each catalytic domain is used once- "noniterative"

Recognized 1991

Type II PKS: Catalytic domains are individual proteins

A Catalytic domain is used for multiple claisen condensations "iterative"

Recognized 1984

Type III PKS: Similar to Type II

No ACP; all substrates are acyl-CoA

Called chalcone synthases and frequently found in plants

Recognized 1999

Iterative Type I PKS: Catalytic domains are linked together on same polypeptide and used iteratively

Recognized in last 2-3 years

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Please see Fig. 1 in *Current Opinion in Chemical Biology* 7 (2003): 285-295.

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DNA of Type II PKS pathway Actinorhodin was sequenced in mid 1980's:

Inspired the search for the genes and was accomplished in 1984 by Hopwood
(this in turn inspired the search for the erythromycin genes)

Although the genes are known it is not completely clear how the enzymes work
(shown on next pages)- Enzymes observed were:

"Minimal" PKS: ACP, KS, KS β chain length factor (CLF)

KR (ketoreductase) --> reduces carbonyl to hydroxyl

CYC (cyclase) like dehydratases

ARO (aromatase) CYC and ARO very similar --> same type of protein

Questions??

No AT domain (loading by either fatty acid AT, by KS or by ACP)

No TE domain

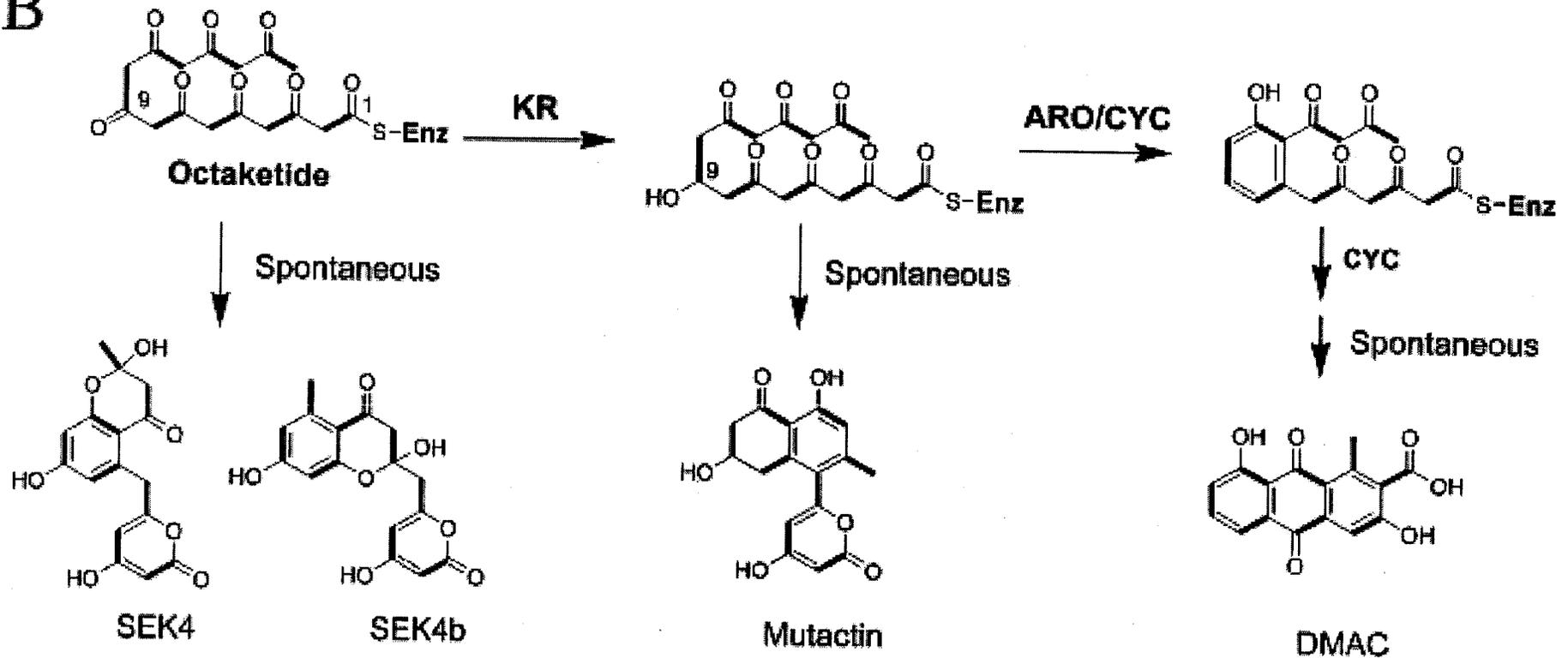
Regioselectivity of reduction/cyclization

Unknown function for second KS domain that lacks active site cysteine (KS β or CLF (Chain Length Factor))

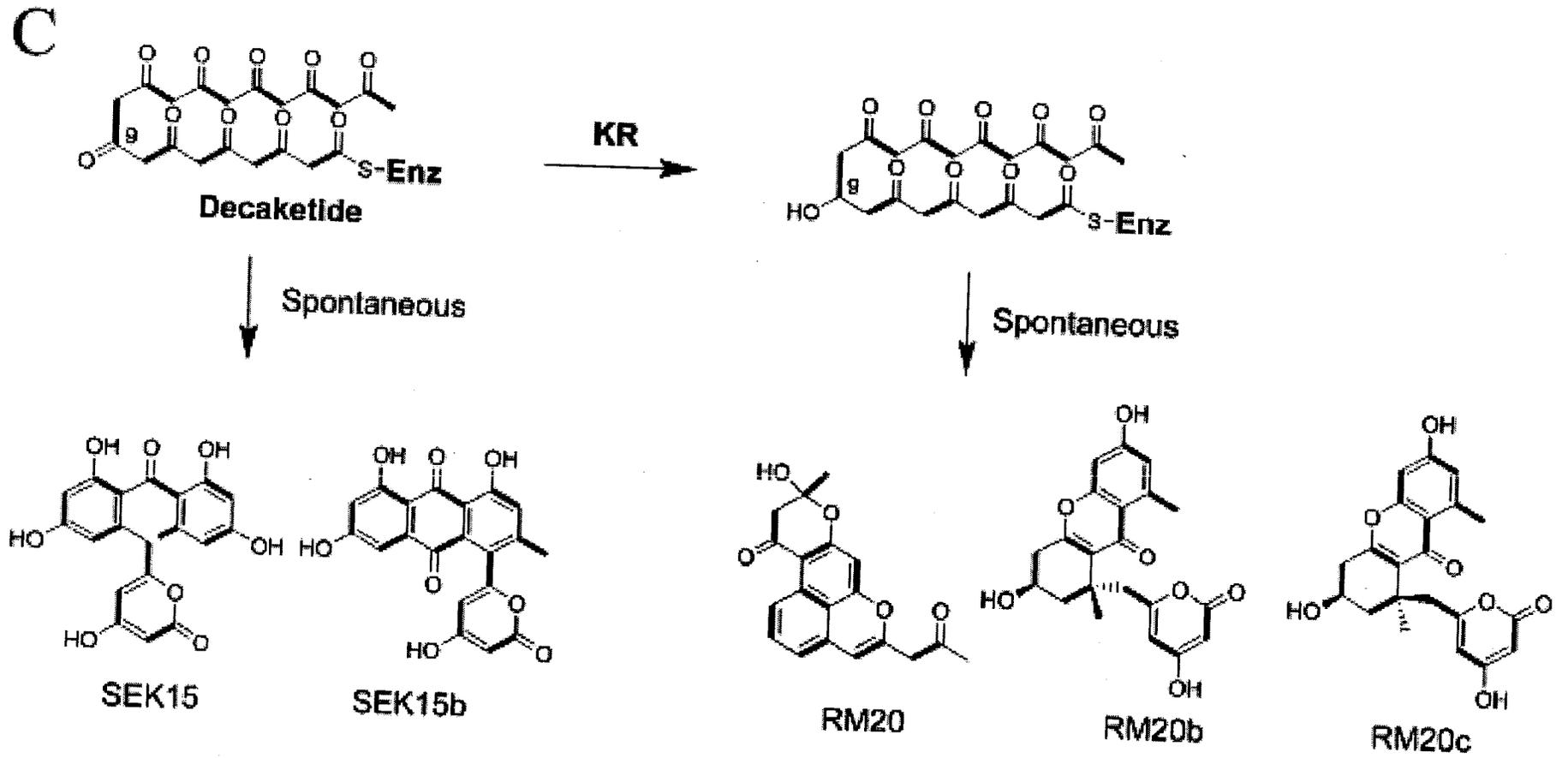
What controls the chain length?

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Please see Fig. 44 in *Nat Prod Rep* 18 (2001): 380-416.

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1. Where is AT domain?

ACP has some endogenous AT activity, though perhaps not enough to support in vivo biosynthesis likely that an AT from FAS synthesis or elsewhere is recruited (Biochemistry (1998) 37, 2084-2088)

2. What controls chain length?

KS forms a heterodimer: second protein called KS β or CLF (Chain Length Factor)
proof that this controls chain length

Crystal structure and mutagenesis studies suggest that a channel formed between the KS and CLF control the length of the polyketide

In vitro: mutate G116T mutation changes from 20 length to >65% C16

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Please see Fig. 1 in *JACS* 125 (2003): 12708-12709.

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Where is the thioesterase?

self cleavage from the thioester (see novobiocin biosynthesis from NRPS section)

A set of "design rules" were proposed in 1995 (Nature (1995) 375, 549-554).
May be outdated

"Minimal PKS" sets the chain length- can switch out an ACP without affecting length but a KS and KSb from same cluster are required for consistent chain lengths K

A KR domain reduces carbonyl to the hydroxyl (act reduces C9)

First ring formation appears to be formed from the minimal PKS (or is not enzymatically catalyzed)

Position of reduction affects regioselectivity of cyclization
reduction at C9 cyclization at C7/C12; if reduce at C7 cyclize at C5/C10
(regiospecificity less well defined when reduction does not occur)

Aromatization domains will only recognize specific carbon length
act ARO recognizes only 16 carbon chains

KR --> each KR has own regioselectivity
--> timing of KR. --> sometimes before ARO/CYC
sometimes acts after ARO/CYC

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Please see Fig. 11 in *Top Curr Chem* 209 (2000): 1.

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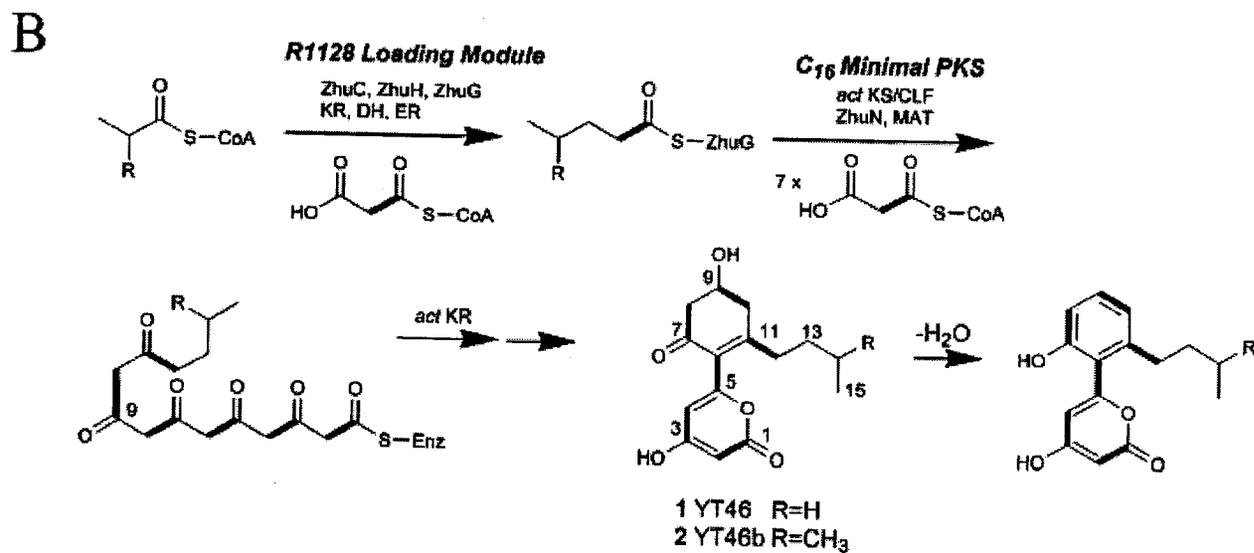
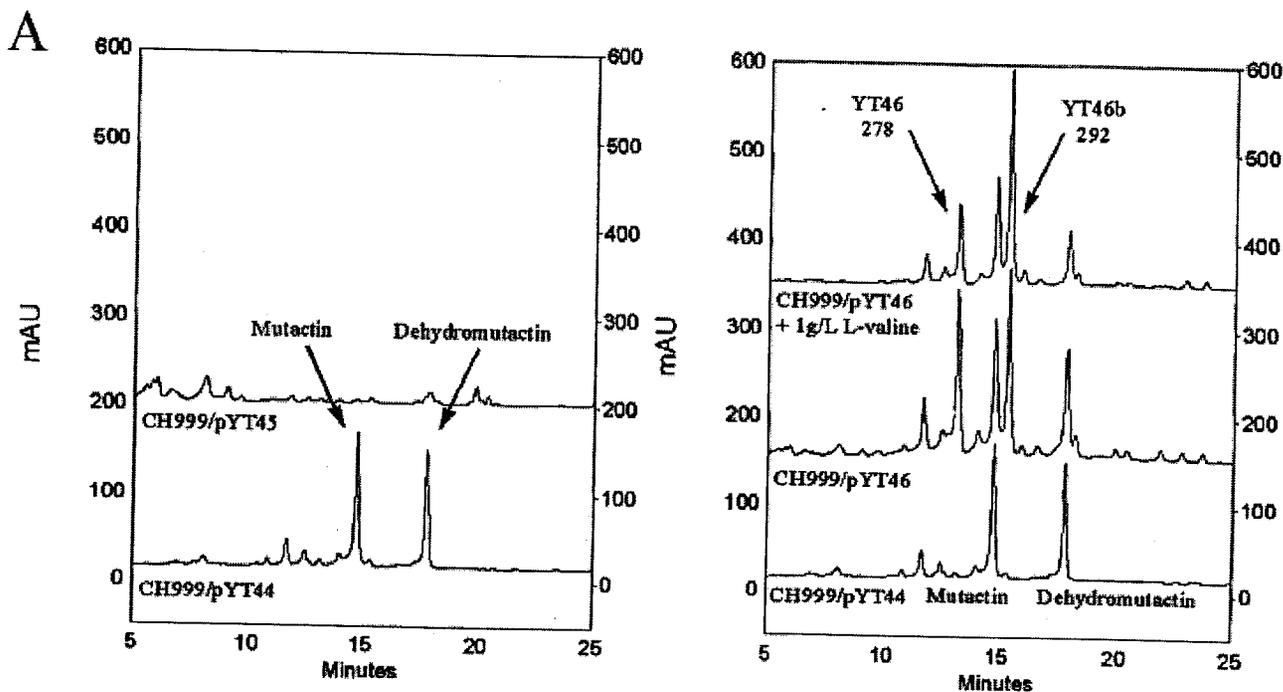


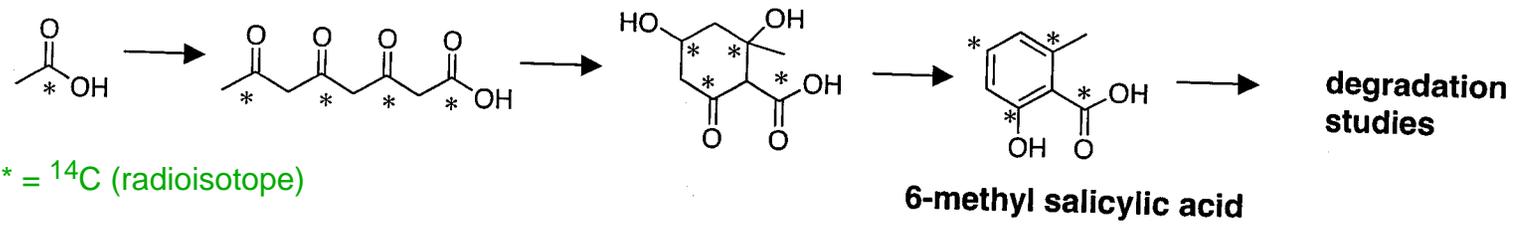
Figure courtesy of:
 Tang, Yi, Taek Soon Lee, and Chaitan Khosla.
 "Engineered Biosynthesis of Regioselectively Modified Aromatic Polyketides
 Using Bimodular Polyketide Synthases." *PLoS Biology* 2, no.2 (2004): 0227-0238.

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Birch (1954) recognized that polyketones can be made by condensation of acetates
What happens if *Penicillium patulum* are fed isotopically labeled acetates?

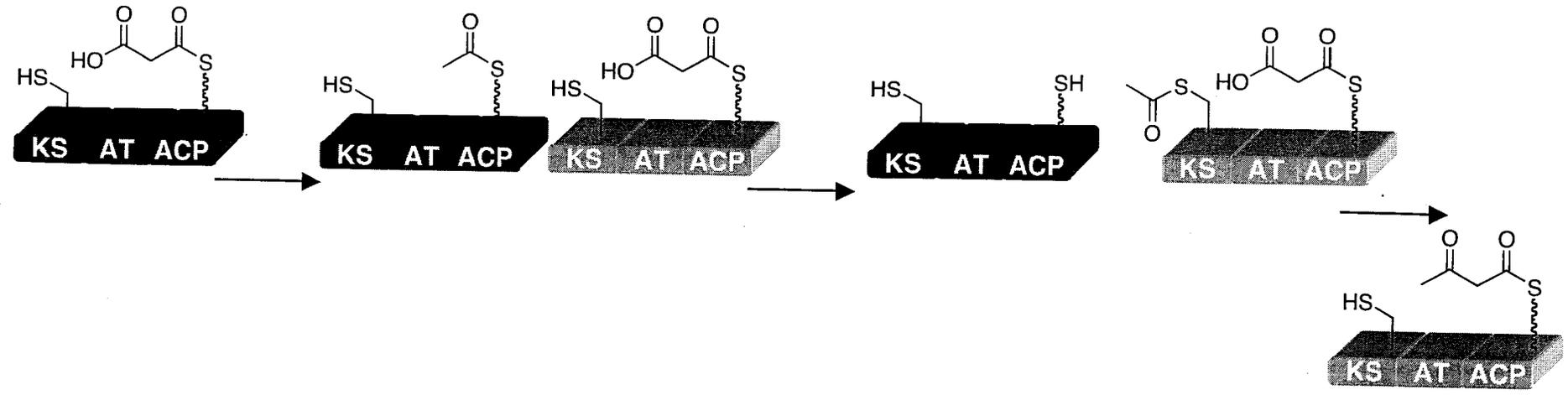


Support for Collie's 1907 hypothesis that aromatic compounds were made from polyketone structures in the cell

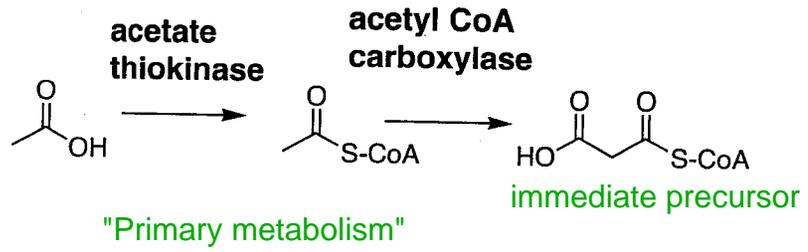
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Labeled acetates can be converted into malonyl-CoA starting material



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Labeling with ^{13}C isotopes facilitated NMR studies

How does labeling work?

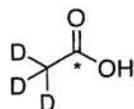
- Indicates the starting material
- Provides structural information

α field signal increases

Assign unlabeled compound- then see which signals increase

Use non- decoupled ^{13}C spectra to observe splitting from the protons

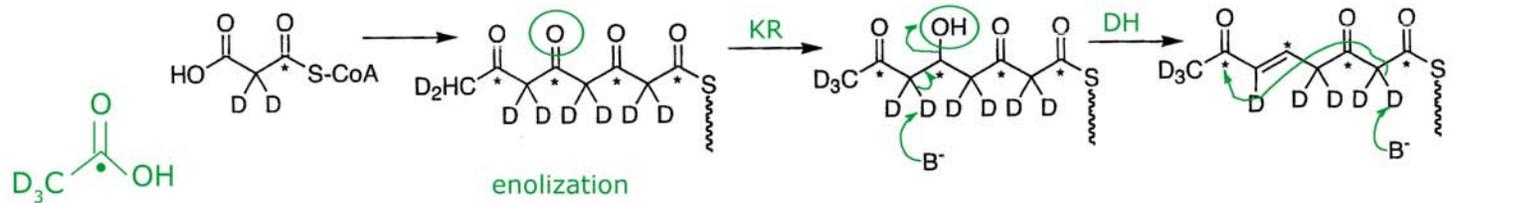
^{13}C , ^2H labeled acetate



upfield shift when a deuterium is attached to a $\text{C}13$ (alpha effect) or is one carbon removed (beta effect)

more pronounced

less pronounced



enolization

incorporation H

"aromatases"

rearomatize

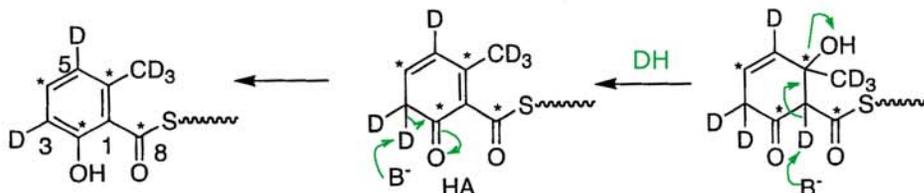
4 enriched signals

→ 8 = singlet

2 = d

4 = d

6 = d



6-methyl salicylic acid

BH A

A = acid, B = base

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Please see Fig. 8 in *Nat Prod Rep* 18 (2001): 380-416.

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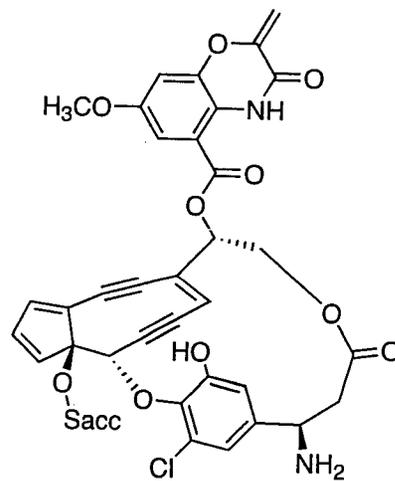
Please see: Mann, John. *Chemical Aspects of Biosynthesis*. New York, NY: Oxford University Press, 1994, pp. 22-23, 26-27.

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Gene clusters of calicheamycin and C-1027 have been shown to be made up of polyketide synthases
Feeding studies in 1989 established an acetate starting material
(Science (2002) 297, 1170, 1173)



C-1027

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Please see Fig. 23 in *Nat Prod Rep* 18 (2001): 380-416.

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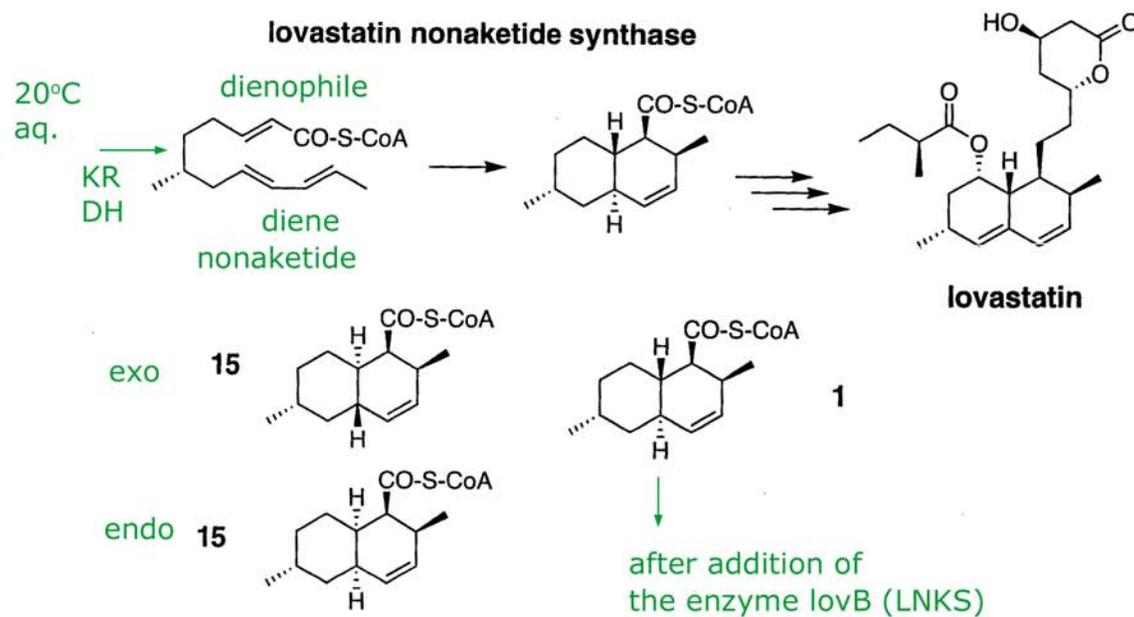
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Diels Alderase?



Has nature discovered the Diels-Alder reaction??



ChemBioChem (2001) 2, 873-875

JACS (2000) 122 11519-11520

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Please see Scheme 1 in *J Biol Chem* 275 (2000): 38393-38401.

Figure removed due to copyright reasons.

Please see Scheme 1 in *ChemBioChem* 4 (2003): 713-715.

Figure removed due to copyright reasons.
Please see Fig. 6-13 in *Nat Prod Rep* 18 (2001): 380-416.